

ab65307

Cathepsin S Activity Assay Kit (Fluorometric)

Instructions for Use

For the rapid, sensitive and accurate measurement of Cathepsin S activity in various samples.

This product is for research use only and is not intended for diagnostic use.

PLEASE NOTE: With the acquisition of BioVision by Abcam, we have made some changes to component names and packaging to better align with our global standards as we work towards environmental-friendly and efficient growth. You are receiving the same high-quality products as always, with no changes to specifications or protocols.

Table of Contents

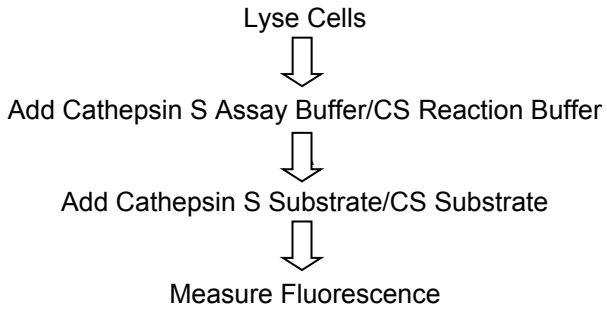
| | |
|---------------------------|---|
| 1. Overview | 3 |
| 2. Protocol Summary | 4 |
| 3. Components and Storage | 5 |
| 4. Assay Protocol | 6 |
| 5. Troubleshooting | 8 |

1. Overview

Apoptosis can be mediated by mechanisms other than the traditional caspase-mediated cleavage cascade. There is growing recognition that alternative proteolytic enzymes such as the lysosomal cathepsin proteases may initiate or propagate proapoptotic signals. Cathepsins are lysosomal enzymes that are also used as sensitive markers in various toxicological investigations.

Abcam's Cathepsin S Activity Assay Kit is a fluorescence-based assay that utilizes the preferred cathepsin-S substrate sequence VVR labeled with AFC (amino-4-trifluoromethyl coumarin). Cell lysates or other samples that contain cathepsin-S will cleave the synthetic substrate VVR-AFC to release free AFC. The released AFC can easily be quantified using a fluorometer or fluorescence plate reader.

2. Protocol Summary



3. Components and Storage

A. Kit Components

| Item | Quantity |
|---|-------------|
| Cathepsin S Lysis Buffer/CS Cell Lysis Buffer | 25 mL |
| Cathepsin S Assay Buffer/CS Reaction Buffer | 5 mL |
| Cathepsin S Substrate/CS Substrate Ac-VVR-AFC (10 mM) | 200 μ L |
| Cathepsin Inhibitor/CS Inhibitor (1mM) | 20 μ L |

* Store kit at -20°C (Store Cathepsin S Lysis Buffer/CS Cell Lysis Buffer and Cathepsin S Assay Buffer/CS Reaction Buffer at 4°C after opening). Protect Cathepsin S Substrate/CS Substrate from light. All reagents are stable for 6 months under proper storage conditions.

B. Additional Materials Required

- Microcentrifuge
- Pipettes and pipette tips
- Fluorometer or fluorescent microplate reader

- 96 well plate
- Orbital shaker

4. Assay Protocol

1. Collect cells ($1-5 \times 10^6$) by centrifugation.

Note:

Use 50-200 μg cell lysates (in 50 μL of Cathepsin S Lysis Buffer/Cell lysis Buffer) if protein concentration has been measured.

2. Lyse cells in 50 μL of chilled Cathepsin S Lysis Buffer/CS Cell Lysis Buffer. Incubate cells on ice for 10 min.

3. Centrifuge at top speed in a microcentrifuge for 5 min, transfer the supernatant to a new tube. Add 50 μL of cell lysate to a 96-well plate.

4. Add 50 μL of Cathepsin S Assay Buffer/CS Reaction Buffer to each sample.

5. Add 2 μL of the 10 mM Cathepsin S Substrate/CS Substrate Ac-VVR-AFC (200 μM final concentration).

Note:

For negative control, add 2 μL of Cathepsin Inhibitor/CS Inhibitor prior to adding Cathepsin S Substrate/CS Substrate, or make a reaction mixture that does not contain sample as control.

6. Incubate at 37°C for 1-2 hours.
7. Read samples in a fluorometer equipped with a 400-nm excitation filter and 505nm emission filter. You may also perform the entire assay directly in a 96-well plate.
8. Fold-increase in Cathepsin S activity can be determined by comparing the relative fluorescence units (RFU) with the level of the uninduced control or the negative control sample. If desired, the units of cathepsin S can be determined by generating a standard curve using free AFC under your assay conditions.

5. Troubleshooting

| Problem | Reason | Solution |
|-------------------|--|---|
| Assay not working | Cells did not lyse completely | Re-suspend the cell pellet in the lysis buffer and incubate as described in the datasheet |
| | Experiment was not performed at optimal time after apoptosis induction | Perform a time-course induction experiment for apoptosis |
| | Plate read at incorrect wavelength | Ensure you are using appropriate reader and filter settings (refer to datasheet) |
| High Background | Increased amount of cell lysate used | Refer to datasheet and use the suggested cell number to prepare lysates |
| | Increased amounts of components added due to incorrect pipetting | Use calibrated pipettes |
| | Incubation of cell samples for extended periods | Refer to datasheet and incubate for exact times |
| | Use of expired kit or improperly stored reagents | Always check the expiry date and store the individual components appropriately |
| | Contaminated cells | Check for bacteria/ yeast/ mycoplasma contamination |

| Problem | Reason | Solution |
|-----------------------|---|---|
| Lower signal levels | Cells did not initiate apoptosis | Determine the time-point for initiation of apoptosis after induction (time-course experiment) |
| | Very few cells used for analysis | Refer to datasheet for appropriate cell number |
| | Use of samples stored for a long time | Use fresh samples or aliquot and store and use within one month for the assay |
| | Incorrect setting of the equipment used to read samples | Refer to datasheet and use the recommended filter setting |
| | Allowing the reagents to sit for extended times on ice | Always thaw and prepare fresh reaction mix before use |
| Unanticipated results | Measured at incorrect wavelength | Check the equipment and the filter setting |
| | Cell samples contain interfering substances | Troubleshoot if it interferes with the kit (run proper controls) |

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| Samples with erratic readings | Uneven number of cells seeded in the wells | Seed only equal number of healthy cells (correct passage number) |
| | Samples prepared in a different buffer | Use the cell lysis buffer provided in the kit |
| | Adherent cells dislodged and lost at the time of experiment | Perform experiment gently and in duplicates/triplicates; apoptotic cells may become floaters |
| | Cell/ tissue samples were not completely homogenized | Use Dounce homogenizer (increase the number of strokes); observe efficiency of lysis under microscope |
| | Samples used after multiple freeze-thaw cycles | Aliquot and freeze samples, if needed to use multiple times |
| | Presence of interfering substance in the sample | Troubleshoot as needed |
| | Use of old or inappropriately stored samples | Use fresh samples or store at correct temperatures until use |

For further technical questions please do not hesitate to contact us by email (technical@abcam.com) or phone (select “*contact us*” on www.abcam.com for the phone number for your region).

Technical Support

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